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# Sample Preparation Protocol SP-02
# Washing Technique for *C. elegans* (nematode)

## Scope

This protocol is the most basic cleaning procedure for use on any culture of *C.elegans* (nematode), prior to running samples on the COPAS instruments. No size selectivity is accomplished using this technique.

## Materials

M9 buffer with 0.01% Triton X-100 (See Sample Preparation Protocol SP-06)
180µm nylon mesh (Millipore Corp. catalog number NY8H04700)
15 ml conical tube
Centrifuge
Microscope
Agar Plates with *C. elegans* population

## Procedure

Wash animals off agar medium using M9 buffer. Use approximately 3 ml of the diluent per plate. In the event of heavy contamination of the worm culture, or if liquid medium is used, the sample should be passed through a 180 µm nylon mesh to remove larger clumps of eggs and debris.

Place the worms into a 15 ml conical tube.

Pellet worms by centrifugation (at low speed) or by settling, and re-suspend in 10 ml of M9 buffer.

Repeat washing step and re-suspend in 10 ml of M9 buffer. Determine the concentration of animals/mL using a microscope.

Adjust the concentration of animals to between 500 and 2000 per ml, by adding diluent buffer.

NOTE: For most applications, two agar plates with moderately heavy growth are enough for approximately 100 ml of final *C. elegans* preparation at a concentration of 1000 organisms / ml.

NOTE: If the preparation is excessively debris ridden, perform a sucrose float technique.

**Questions?**
**For further information, please contact Union Biometrica, Inc. directly at 617.591.1211 or email your questions to appsupport@unionbio.com**